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Mini review

POTENTIAL ROLES OF THE NF κ B AND GLUTATHIONE PATHWAYS IN MATURE HUMAN ERYTHROCYTES [#]

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Abstract: Anucleated erythrocytes were long considered as oxygen-transporting cells with limited regulatory functions. Components of different nuclear signaling pathways have not been investigated in those cells, yet. Surprisingly, we repeatedly found significant amounts of transcription factors in purified erythrocyte preparations, i.e. nuclear factor κ B (NF κ B), and major components of the canonical NF κ B signaling pathway. To investigate the functional role of NF κ B signaling, the effects of the preclinical compounds Bay 11-7082 and parthenolide on the survival of highly purified erythrocytes were investigated. Interestingly, both inhibitors of the NF κ B pathway triggered erythrocyte programmed cell death as demonstrated by enhanced phospholipid scrambling

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Abbreviations used: Ca²⁺ – calcium; ERK2 – extracellular signal-regulated kinase 2; GPIIb/IIIa – glycoprotein IIb/IIIa; GSH – reduced glutathione; ICAM-1 – inter-cellular adhesion molecule; IFN- γ – interferon- γ ; I κ B- α – inhibitor of kappaB; IKK- α – I κ B kinase- α ; IL-6 – interleukin-6; IL-8 – interleukin-8; JNK1 – c-Jun N-terminal kinase 1; NF κ B – nuclear factor κ B; PS – phosphatidylserine; R – putative phosphatidylserine receptor

(phosphatidylserine exposure) and cell shrinkage. Anucleated erythrocytes are an ideal cellular model allowing the study of nongenomic mechanisms contributing to suicidal cell death. As NF κ B inhibitors might also interfere with the anti-oxidative defense systems of the cell, we measured the levels of reduced glutathione (GSH) after challenge with the inhibitors. Indeed, incubation of erythrocytes with Bay 11-7082 clearly decreased erythrocyte GSH levels. In conclusion, the pharmacological inhibitors of the NF κ B pathway Bay 11-7082 and parthenolide interfere with the survival of erythrocytes involving mechanisms other than disruption of NF κ B-dependent gene expression. Besides affecting erythrocyte survival, NF κ B inhibition and induction of erythrocyte phosphatidylserine exposure may influence blood clotting. Future studies will be aimed at discriminating between NF κ B-dependent and NF κ B-independent GSH-mediated effects of Bay 11-7082 and parthenolide on erythrocyte death.

Key words: Eryptosis, Nuclear factor kappa B, NF κ B, IKK- α , I κ B- α , Bay 11-7082, Parthenolide

EXPRESSION OF DIFFERENT COMPONENTS OF THE CANONICAL NF κ B PATHWAY IN MATURE HUMAN ERYTHROCYTES

NF κ B was first discovered and characterized 25 years ago in immune cells [1]. The predominant inactive form of cytoplasmic NF κ B is a heterotrimeric protein consisting of RelA (p65), p50 and the inhibitor of κ B (I κ B)- α subunit. Upon cellular stimulation via different surface receptors of the innate immune system as well as receptors for cytokines, e.g. Toll-like-receptors (TLRs), B cell receptor (BCR), T cell receptor (TCR) or tumor necrosis factor receptor (TNFR), I κ B- α is phosphorylated on serine 32 and 36 by I κ B kinase (IKK), followed by polyubiquitination and 26S proteasomal degradation. Activated NF κ B then translocates to the nucleus as a p65-p50 heterodimer, where it acts as a transcription factor, thereby enhancing the expression of a variety of genes involved in the regulation of immune responses. NF κ B signaling thus leads to the induction of various cytokines (IL-6, IL-8, IFN- γ), adhesion molecules (ICAM1), acute phase response proteins (complement factor B) and regulator proteins of cell survival. Taken together, NF κ B plays an important role in a great variety of immune reactions [2-5].

Mature human erythrocytes are devoid of nuclei, and nuclear signaling should not play a functional role in this specialized cell type. However, we could repeatedly demonstrate by Western blotting that purified human erythrocyte concentrates from various volunteers expressed both subunits of active NF κ B, namely RelA and p50 (Fig. 1). Further purification and dilution experiments clearly showed that the immuno-reactive bands did not originate from contaminating white blood cells or thrombocytes [6]. Mature erythrocytes expressed not only active NF κ B but also the regulatory upstream elements of the NF κ B signaling pathway I κ B- α kinase (IKK) and I κ B- α (Fig. 1). Taking into

account that the canonical pathway of NF κ B activation, including the expression levels of its components, is tightly regulated, these results were quite astonishing. Recently, experimental evidence was provided that mature erythrocytes contain functional 20S proteasomes [7]. Due to proteasomal degradation, the halflife of I κ B- α should thus be relatively short. However, the finding that I κ B- α is expressed in mature human erythrocytes, which have an average life span of approx. 120 days, points to special mechanisms of protein stabilization.

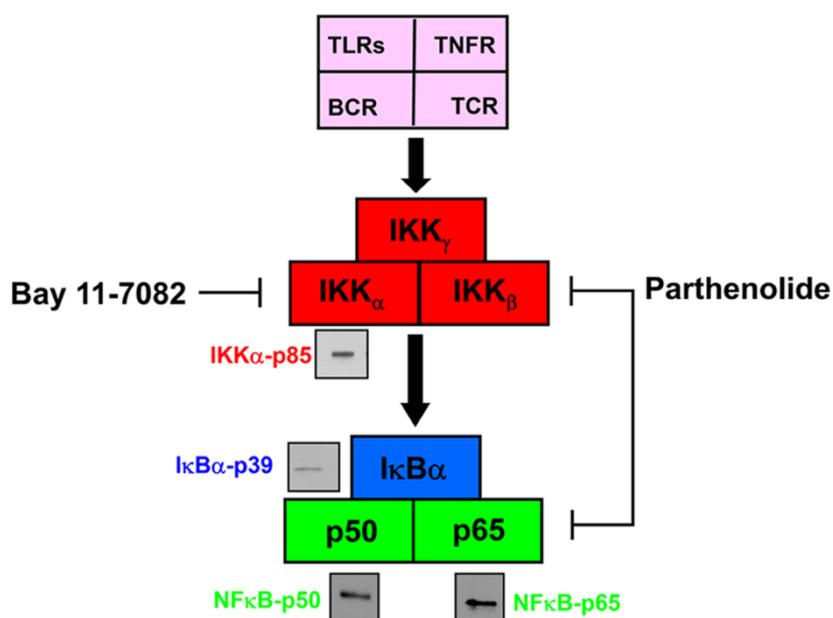


Fig. 1. Expression of components of the canonical NF κ B signaling pathway in mature human erythrocytes. The scheme depicts the elements of the canonical NF κ B signaling pathway including the upstream regulatory components. In addition, the expression of both subunits of active NF κ B, namely RelA and p50, as well as IKK and I κ B- α in mature human erythrocytes is shown. For this, erythrocyte extracts were subjected to Western blotting, and IKK1- α (85 kD), I κ B- α (39 kD), RelA (65 kD) and the NF κ B subunit p50 (50 kD) were detected using specific antibodies. Abbreviations: BCR – B cell receptor; I κ B α – inhibitor of kappa B; IKK – I κ B α kinase; p50 – 50 kD subunit of NF κ B; RelA – 65 kD subunit of NF κ B; TCR – T cell receptor, TLR – toll-like receptor; TNFR – tumor necrosis factor receptor.

The fact that mature erythrocytes still contain detectable amounts of the components of the NF κ B pathway does not imply that the pathway is still functional. These “leftover” proteins might also represent functionally irrelevant remnants. To investigate whether NF κ B plays a functional role in mature erythrocytes, we therefore tested different NF κ B inhibitors in our experimental system.

TARGETING OF THE NF κ B AND GLUTATHIONE PATHWAYS BY BAY 11-7082 AND PARTHENOLIDE

Since NF κ B signaling is crucial for the activation and regulation of various immune responses, the different subunits of the protein and the other regulators of the NF κ B signaling pathway were immediately recognized as important drug targets. Indeed, there are hundreds of inhibitors available to block NF κ B activity [8]. From these inhibitors, we chose two chemically unrelated candidates that selectively block at different sites of the NF κ B pathway (Fig. 1 and Tab. 1). The synthetic and lipophilic (E)-3-(4-methylphenylsulfonyl)-2-propenenitrile (Bay 11-7082) blocks the kinase activity of IKK, thereby preventing I κ B- α phosphorylation and degradation [9], and the sesquiterpene lactone (SL) and active component of the medical herb feverfew (*Tanacetum parthenium*) parthenolide blocks IKK α [10] and the NF κ B subunit p65 (RelA) [11, 12]. In addition, it could be shown that a methylene-carbonyl substructure is crucial for SL-based inhibition of RelA at cysteine 38 [13].

Tab. 1. Targeting of the NF κ B pathway in erythrocytes by specific inhibitors

Inhibitor	Target(s)	EC ₅₀ PS exposure	EC ₅₀ Cell shrinkage	Intracellular Ca ²⁺ level ^d
Bay 11-7082 ^a	IKK	11 μ M ^c	11 μ M ^c	28% ^{c,d}
Parthenolide ^b	IKK and RelA	30 μ M ^c	35 μ M ^c	19% ^{c,d}
Bay 11-7082 ^c	GSH	EC ₅₀ GSH depletion 2 μ M ^c	EC ₁₀₀ GSH depletion \geq 7.5 μ M ^c	

^a[9], ^b[10, 11], ^c[6], ^dCa²⁺ level is expressed as the percentage of maximum ionomycin-induced Ca²⁺ increase

We treated erythrocytes with pharmacological doses of these inhibitors and found that both inhibitors were very effective inducers of programmed erythrocyte death, as characterized by phosphatidylserine (PS) exposure, cell shrinkage and elevation of intracellular Ca²⁺ (Tab. 1). PS exposure and cell shrinkage are the hallmarks of eryptosis, the suicidal death of erythrocytes [14-16]. In control experiments, we excluded a non-specific lytic effect of the NF κ B inhibitors at the concentrations used [6]. Thus, these results argued in favor of a functional role of NF κ B signaling in erythrocyte survival.

The effects of Bay 11-7082 and parthenolide on erythrocyte programmed cell death described in the previous paragraph were induced by pharmacological concentrations of the inhibitors. Thus, it seems reasonable that the inhibitors exert their effects on erythrocytes by inhibition of a currently unknown function of NF κ B. Nevertheless, at least for parthenolide, it is already known that this drug might also influence the anti-oxidative defense of cells by lowering the level of reduced glutathione (GSH) [17-19]. In a chemical reaction, GSH is added to the double bond of parthenolide, resulting in a parthenolide-GSH-adduct (Fig. 2A). GSH depletion has previously been shown to trigger eryptosis [6].

Theoretical considerations and the comparison of the chemical structure of Bay 11-7082 with the structure of parthenolide led us to the speculation that Bay 11-7082 might also react with GSH to form Bay 11-7082-GSH adducts (Fig. 2B). As in the case of parthenolide, this reaction should decrease the level of GSH in erythrocytes. Indeed, treatment of erythrocytes with Bay 11-7082 completely depleted GSH from the cells with an IC₅₀ of approximately 2 μM [6]. Taken together, the NFκB pathway inhibitors Bay 11-7082 and parthenolide induce erythrocyte cell death by simultaneously targeting NFκB signaling and the anti-oxidative defense system of the cells (Fig. 2C). Thus, the link between inhibitor-induced eryptosis and the presence of the NFκB pathway in mature human erythrocytes is elusive and needs further investigations. To clearly demonstrate an unconventional nongenomic role of NFκB in erythrocytes, future studies using more direct genetic approaches with targeted modifications of individual components of the NFκB pathway are needed. For instance, the effects of Bay 11-7082 and parthenolide on erythrocytes from IKK- [20] or p50/NFκB-deficient mice [21] could be analyzed.

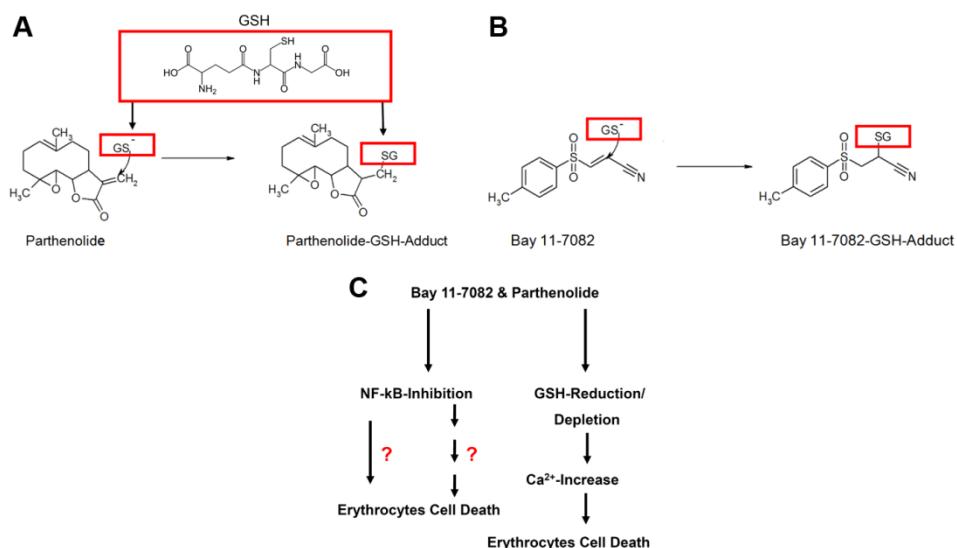


Fig. 2. Targeting of glutathione-dependent anti-oxidative defense by NFκB inhibitors. A – Reduced glutathione might chemically react with parthenolide (nucleophilic addition to a double bond) to form parthenolide-GSH adducts, thereby lowering the GSH level of cells [17]. B – Bay 11-7082 also contains a double bond which might be susceptible to a nucleophilic attack. This reaction would likewise lead to the formation of Bay 11-7082-GSH adducts and depletion of GSH in the cell. C – The scheme depicts NFκB signaling and GSH depletion as the main targets of Bay 11-7082 and parthenolide in erythrocytes. Interference with both pathways finally increases Ca²⁺ and leads to programmed erythrocyte death. GSH, reduced glutathione; NFκB, nuclear factor κB.

It should be kept in mind that there may be a direct interaction between the oxidative status of the erythrocytes and the expression levels of the members of the NF κ B pathway. Recent results from our laboratory showing a correlation between GSH and NF κ B expression levels in erythrocytes with different age support this assumption (data not shown).

POSSIBLE ROLE OF THE NF κ B AND THE GLUTATHIONE PATHWAY IN BLOOD CLOTTING

NF κ B is expressed in nucleus-free erythrocytes [6] and thrombocytes [22]. Thrombocytes play an essential role in hemostasis (blood clotting in response to injury) and in pathological conditions such as inflammation, immunity and tumor progression [23]. Recent observations revealed that the pharmacological drugs Bay 11-7082 and parthenolide also affect the function of thrombocytes [24].

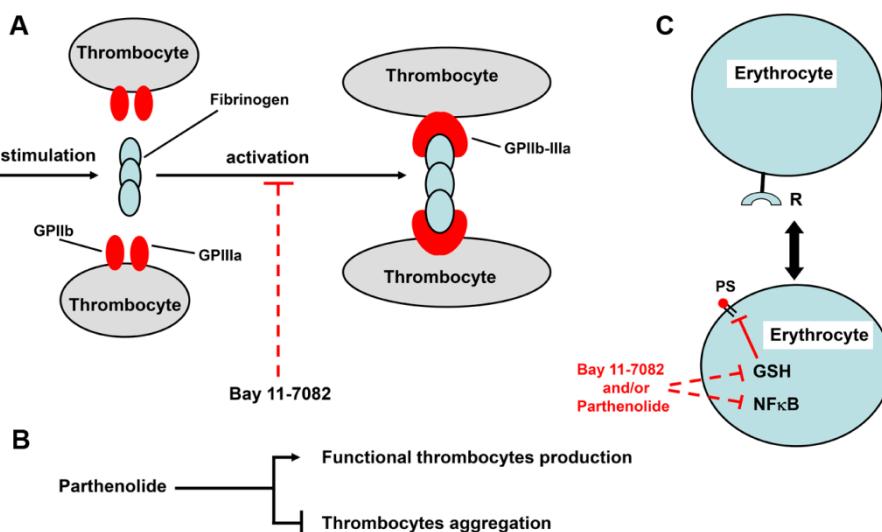


Fig. 3. Effects of NF κ B inhibitors on adhesion of thrombocytes and erythrocytes. A – After stimulation and activation, thrombocytes aggregate via fibrinogen-mediated glycoprotein IIb/IIIa interaction (modified according to Dangel, [35]). Anti-thrombocyte agents, such as Bay 11-7082, inhibit this activation, thereby preventing thrombocyte aggregation [24]. B – Parthenolide is able to stimulate functional platelet production by inhibition of NF κ B signaling [25] and to reduce thrombocyte aggregation [26]. C – Bay 11-7082 and parthenolide also inhibit the anti-oxidative defense and/or NF κ B signaling in erythrocytes and concentration-dependently induce phosphatidylserine (PS) exposure on the erythrocyte surface. PS on the cellular surface then interacts with its putative receptor on neighboring erythrocytes (see double back arrow), and leads to intercellular adhesion of erythrocytes. Abbreviations: GPIIb-IIIa – glycoprotein IIb/IIIa; GSH – reduced glutathione; NF κ B – nuclear factor κ B; PS – phosphatidylserine; R – putative phosphatidylserine receptor.

As illustrated in Fig. 3A, Bay 11-7082 is able to reduce fibrinogen binding to glycoprotein GPIIb-IIIa (also named α IIb β 3 according to integrin nomenclature), thereby preventing thrombocyte aggregation. In addition, Bay 11-7082 inhibits thrombocyte activation and aggregation, and forces them to retain their granular contents, effects that are mediated by inhibition of JNK1 and ERK2 phosphorylation [25]. The fact that fibrinogen positively regulates NF κ B activation and expression of inflammatory chemokines in endothelial cells [26] shows the multifunctional importance and applicability of such anti-inflammatory drugs as Bay 11-7082. Recently, it was shown that the electrophilic and anti-oxidant compound parthenolide stimulates functional thrombocyte production by inhibition of NF κ B signaling in megakaryocytes [27]. Parthenolide is also able to inhibit thrombocyte aggregation [28] (Fig. 3B), and it was reported more than three decades ago that depletion of reduced glutathione (GSH) by the thiol oxidizing agent “diamide” resulted in immediate inhibition of thrombocyte aggregation and clot retraction [29]. These observations point to an important role of GSH in thrombocyte function. In line with this, Bay 11-7082 and parthenolide, with their capabilities to deplete reduced glutathione (GSH) and to inhibit NF κ B activity [6], could control thrombocyte functions and blood clotting, and inhibit the risk of thrombosis. On the other hand, Bay 11-7082 and parthenolide also target NF κ B and GSH in erythrocytes, and the challenge of erythrocytes with the inhibitors leads to increased PS exposure on the outer leaflet of the plasma membrane. PS then binds to its receptor [30] expressed on adjacent erythrocytes or on other cell types, such as macrophages or endothelial cells (Fig. 3C). Thus, increased adherence of erythrocytes might enhance the stabilization of an already formed blood clot [31, 32] or even result in blood clotting in some special circumstances, for example during sickle cell crisis [33], infection with plasmodia [34] or other pathological conditions. It is therefore crucial to analyze the concentration-dependent effects of Bay 11-7082 and parthenolide on thrombocyte aggregation to assess the minimal effective dose at which thrombocytes aggregation is inhibited by reduction of their adhesion molecules. As previously shown, low dose inhibitor concentrations ($\leq 5 \mu\text{M}$ for Bay 11-7082 and $\leq 10 \mu\text{M}$ for parthenolide) do not lead to erythrocyte cell death [6]. Due to the very effective depletion of GSH by Bay 11-7082 ($\text{IC}_{50} = 2 \mu\text{M}$; see also Tab. 1), it is tempting to speculate that low concentrations of the substance may reduce the expression levels of adhesion molecules on the thrombocyte surface without leading to cell death.

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REFERENCES

1. Sen, R. and Baltimore, D. Multiple nuclear factors interact with the immunoglobulin enhancer sequences. *Cell* **46** (1986) 705-716.
2. Ghosh, S. and Karin, M. Missing pieces in the NF-kappaB puzzle. *Cell* **109** Suppl (2002) S81-S96.
3. Hayden, M.S. and Ghosh, S. Signaling to NF-kappaB. *Genes Dev.* **18** (2004) 2195-2224.
4. Hayden, M.S. and Ghosh, S. NF-kappaB in immunobiology. *Cell. Res.* **21** (2011) 223-244.
5. Sun, S.C. Non-canonical NF-kappaB signaling pathway. *Cell. Res.* **21** (2011) 71-85.
6. Ghashghaeinia, M., Toulany, M., Saki, M., Bobbala, D., Fehrenbacher, B., Rupec, R., Rodemann, H.P., Ghoreschi, K., Rocken, M., Schaller, M., Lang, F. and Wieder, T. The NFkB pathway inhibitors Bay 11-7082 and parthenolide induce programmed cell death in anucleated Erythrocytes. *Cell. Physiol. Biochem.* **27** (2011) 45-54.
7. Neelam, S., Kakhiashvili, D.G., Wilkens, S., Levene, S.D. and Goodman, S.R. Functional 20S proteasomes in mature human red blood cells. *Exp. Biol. Med.* (Maywood.) **236** (2011) 580-591.
8. Gilmore, T.D. and Herscovitch, M. Inhibitors of NF-kappaB signaling: 785 and counting. *Oncogene* **25** (2006) 6887-6899.
9. Pierce, J.W., Schoenleber, R., Jesmok, G., Best, J., Moore, S.A., Collins, T. and Gerritsen, M.E. Novel inhibitors of cytokine-induced IkappaBalphaphosphorylation and endothelial cell adhesion molecule expression show anti-inflammatory effects in vivo. *J. Biol. Chem.* **272** (1997) 21096-21103.
10. Kwok, B.H., Koh, B., Ndubuisi, M.I., Elofsson, M. and Crews, C.M. The anti-inflammatory natural product parthenolide from the medicinal herb Feverfew directly binds to and inhibits IkappaB kinase. *Chem. Biol.* **8** (2001) 759-766.
11. Garcia-Pineres, A.J., Castro, V., Mora, G., Schmid, T.J., Strunck, E., Pahl, H.L. and Merfort, I. Cysteine 38 in p65/NF-kappaB plays a crucial role in DNA binding inhibition by sesquiterpene lactones. *J. Biol. Chem.* **276** (2001) 39713-39720.
12. Garcia-Pineres, A.J., Lindenmeyer, M.T. and Merfort, I. Role of cysteine residues of p65/NF-kappaB on the inhibition by the sesquiterpene lactone parthenolide and N-ethyl maleimide, and on its transactivating potential. *Life Sci.* **75** (2004) 841-856.
13. Wagner, S., Hofmann, A., Siedle, B., Terfloth, L., Merfort, I. and Gasteiger, J. Development of a structural model for NF-kappaB inhibition of sesquiterpene lactones using self-organizing neural networks. *J. Med. Chem.* **49** (2006) 2241-2252.

14. Foller, M., Bobbala, D., Koka, S., Huber, S.M., Gulbins, E. and Lang, F. Suicide for survival--death of infected erythrocytes as a host mechanism to survive malaria. *Cell. Physiol. Biochem.* **24** (2009) 133-140.
15. Lang, F., Lang, K.S., Lang, P.A., Huber, S.M. and Wieder, T. Mechanisms and significance of eryptosis. *Antioxid. Redox. Signal.* **8** (2006) 1183-1192.
16. Lang, F., Gulbins, E., Lang, P.A., Zappulla, D. and Foller, M. Ceramide in suicidal death of erythrocytes. *Cell. Physiol. Biochem.* **26** (2010) 21-28.
17. Koprowska, K. and Czyz, M. Molecular mechanisms of parthenolide's action: Old drug with a new face. *Postepy Hig. Med. Dosw.* **64** (2010) 100-114.
18. Wen, J., You, K.R., Lee, S.Y., Song, C.H. and Kim, D.G. Oxidative stress-mediated apoptosis. The anticancer effect of the sesquiterpene lactone parthenolide. *J. Biol. Chem.* **277** (2002) 38954-38964.
19. Zhang, S., Ong, C.N. and Shen, H.M. Critical roles of intracellular thiols and calcium in parthenolide-induced apoptosis in human colorectal cancer cells. *Cancer Lett.* **208** (2004) 143-153.
20. Pasparakis, M., Luedde, T. and Schmidt-Suprian, M. Dissection of the NF-kappaB signalling cascade in transgenic and knockout mice. *Cell Death Differ.* **13** (2006) 861-872.
21. Snapper, C.M., Zelazowski, P., Rosas, F.R., Kehry, M.R., Tian, M., Baltimore, D. and Sha, W.C. B cells from p50/NF-kappa B knockout mice have selective defects in proliferation, differentiation, germ-line CH transcription, and Ig class switching. *J. Immunol.* **156** (1996) 183-191.
22. Spinelli, S.L., Casey, A.E., Pollock, S.J., Gertz, J.M., McMillan, D.H., Narasipura, S.D., Mody, N.A., King, M.R., Maggirwar, S.B., Francis, C.W., Taubman, M.B., Blumberg, N. and Phipps, R.P. Platelets and megakaryocytes contain functional nuclear factor-kappaB. *Arterioscler. Thromb. Vasc. Biol.* **30** (2010) 591-598.
23. Morel, O., Jesel, L., Freyssinet, J.M. and Toti, F. Cellular mechanisms underlying the formation of circulating microparticles. *Arterioscler. Thromb. Vasc. Biol.* **31** (2011) 15-26.
24. Malaver, E., Romaniuk, M.A., D'Atri, L.P., Pozner, R.G., Negrotto, S., Benzadon, R. and Schattner, M. NF-kappaB inhibitors impair platelet activation responses. *J. Thromb. Haemost.* **7** (2009) 1333-1343.
25. Lee, H.S., Kim, S.D., Lee, W.M., Endale, M., Kamruzzaman, S.M., Oh, W.J., Cho, J.Y., Kim, S.K., Cho, H.J., Park, H.J. and Rhee, M.H. A noble function of BAY 11-7082: Inhibition of platelet aggregation mediated by an elevated cAMP-induced VASP, and decreased ERK2/JNK1 phosphorylations. *Eur. J. Pharmacol.* **627** (2010) 85-91.
26. Guo, M., Sahni, S.K., Sahni, A. and Francis, C.W. Fibrinogen regulates the expression of inflammatory chemokines through NF-kappaB activation of endothelial cells. *Thromb. Haemost.* **92** (2004) 858-866.
27. Sahler, J., Bernard, J.J., Spinelli, S.L., Blumberg, N. and Phipps, R.P. The Feverfew plant-derived compound, parthenolide enhances platelet

- production and attenuates platelet activation through NF-kappaB inhibition. **Thromb. Res.** 127 (2011) 426-434.
- 28. Groenewegen, W.A. and Heptinstall, S. A comparison of the effects of an extract of feverfew and parthenolide, a component of feverfew, on human platelet activity in-vitro. **J. Pharm. Pharmacol.** 42 (1990) 553-557.
 - 29. Matsuda, S., Ikeda, Y., Aoki, M., Toyama, K., Watanabe, K. and Ando, Y. Role of reduced glutathione on platelet functions. **Thromb. Haemost.** 42 (1979) 1324-1331.
 - 30. Fadok, V.A., Bratton, D.L., Rose, D.M., Pearson, A., Ezekowitz, R.A. and Henson, P.M. A receptor for phosphatidylserine-specific clearance of apoptotic cells. **Nature** 405 (2000) 85-90.
 - 31. Steffen, P., Jung, A., Nguyen, D.B., Muller, T., Bernhardt, I., Kaestner, L. and Wagner, C. Stimulation of human red blood cells leads to Ca(2+)-mediated intercellular adhesion. **Cell. Calcium** 50 (2011) 54-61.
 - 32. Kaestner, L., Tabellion, W., Lipp, P. and Bernhardt, I. Prostaglandin E2 activates channel-mediated calcium entry in human erythrocytes: an indication for a blood clot formation supporting process. **Thromb. Haemost.** 92 (2004) 1269-1272.
 - 33. Lang, K.S., Roll, B., Myssina, S., Schittenhelm, M., Scheel-Walter, H.G., Kanz, L., Fritz, J., Lang, F., Huber, S.M. and Wieder, T. Enhanced erythrocyte apoptosis in sickle cell anemia, thalassemia and glucose-6-phosphate dehydrogenase deficiency. **Cell. Physiol. Biochem.** 12 (2002) 365-372.
 - 34. Lang, P.A., Kasinathan, R.S., Brand, V.B., Duranton, C., Lang, C., Koka, S., Shumilina, E., Kempe, D.S., Tanneur, V., Akel, A., Lang, K.S., Foller, M., Kun, J.F., Kremsner, P.G., Wesselborg, S., Laufer, S., Clemen, C.S., Herr, C., Noegel, A.A., Wieder, T., Gulbins, E., Lang, F. and Huber, S.M. Accelerated clearance of Plasmodium-infected erythrocytes in sickle cell trait and annexin-A7 deficiency. **Cell. Physiol. Biochem.** 24 (2009) 415-428.
 - 35. Dangel, O. [Wirkung von Stickstoffmonoxid auf die Thrombozytenfunktion von Guanylyl-Cyclase defizienten Mäusen. Lehrstuhl für Pharmakologie und Toxikologie der Fakultät für Medizin] Bochum (2007) 24.