

SHORT REPORT

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Apolipoprotein E polymorphism and the risk of aneurysmal subarachnoid hemorrhage in a South Indian population

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Abstract

Background: The rupture of a brain aneurysm causes bleeding in the subarachnoid space. This is known as aneurysmal subarachnoid hemorrhage (aSAH). We evaluated the association of apolipoprotein E (*APOE*) polymorphism and the risk of aSAH in a South Indian population.

Methods: The study was performed on 200 subjects with aSAH and 253 healthy control subjects. Blood samples (5 ml) were used to isolate DNA and genotyping was performed for rs7412 and rs429358 using a Taqman allelic discrimination assay. Statistical software R.3.0.11 was used to statistically analyze the data and a p value < 0.05 was considered as statistically significant.

Results: We found a significant association with the risk of aSAH in $\epsilon 3/\epsilon 4$ genetic model (OR = 1.91, 95% CI = 1.16–3.14, $p = 0.01$). However, in the other genetic models and allele frequency, there was no significant association with the risk of aSAH. In subtyping, we found a significant association of $\epsilon 2$ allele frequency with posterior communicating artery (PCOM) aneurysm (OR = 3.59, 95% CI = 1.11–11.64, $p = 0.03$).

Conclusion: Our results suggest that *APOE* polymorphism has an influence on the risk of aSAH in this South Indian population, specifically in the PCOM subtype.

Keywords: Aneurysmal subarachnoid hemorrhage, Apolipoprotein E, Polymorphism, Posterior communicating artery aneurysm

Introduction

Aneurysmal subarachnoid hemorrhage (aSAH) is a life-threatening condition that accounts for 5% of strokes [1]. aSAH occurs due to the rupture of an aneurysm that is commonly found in the Circle of Willis region in the brain [2]. A recent report suggested that the overall incidence of aSAH was approximately 9 in 100,000 people per year, but that this can vary significantly with geographical region [3]. In India, the incidence rate was determined as 21.8 in 100,000 people per year [4]. Japan and Finland have higher incidences of aSAH – 22 and 19.7 in 100,000, respectively [5]. Lower incidence rates were reported for South and Central America [3].

Genetic and environmental risk factors play a key role in the formation and rupture of aneurysms. Genome-wide association studies of aSAH in different population have identified many new risk loci associated with aSAH [6].

Apolipoprotein E (*APOE*) is a 299-amino acid protein mainly synthesized by the liver and macrophages [7]. In human beings, it is located in the long arm of chromosome 19 and the protein is encoded by exon 4 [8]. In the brain, it is synthesized by astrocytes and helps in the transport of cholesterol to neurons [9]. *APOE* also helps in neuronal repair [10], cerebral glucose metabolism [11], and differentiation and migration of neurons [12]. The *APOE* gene is polymorphic with three different alleles ($\epsilon 2$, $\epsilon 3$, $\epsilon 4$) and six different genotypes [13]. The combination of polymorphisms of rs7412 and rs429358 will determine the six possible genotypes [14].

In the Asian population, the allele frequencies for $\epsilon 2$, $\epsilon 3$ and $\epsilon 4$ were 5%, 86% and 9%, respectively. The most common genotype in the Asian population is $\epsilon 3/\epsilon 3$ (74%), followed by $\epsilon 3/\epsilon 4$ (16%), $\epsilon 2/\epsilon 4$ (13%), $\epsilon 2/\epsilon 3$ (8.5%), $\epsilon 4/\epsilon 4$ (0.7%) and $\epsilon 2/\epsilon 2$ (0.1%) [15]. The frequencies of *APOE* alleles do not vary with gender [16]. The $\epsilon 4$ allele is associated with unfavorable outcomes after traumatic brain injury [17, 18] and aSAH in the Chinese and Italian population [19, 20]. We performed a meta-analysis that suggested that there is significant association with the risk of aSAH in $\epsilon 2/\epsilon 2$ vs. $\epsilon 3/\epsilon 3$, $\epsilon 2/\epsilon 3$ vs. $\epsilon 3/\epsilon 3$ and $\epsilon 2$ vs. $\epsilon 3$ genetic models and $\epsilon 2$ allele frequency [21].

There is no study that has addressed the relationship between *APOE* polymorphism and the risk of developing aSAH in the Indian population. The aim of this study is to investigate the association of *APOE* polymorphism and aSAH in a South Indian population.

Materials and methods

Study population

The subjects were 200 patients with aneurysmal subarachnoid hemorrhage recruited from the Department of Neurosurgery, NIMHNAS, Bangalore, India, and 253 ethnicity-, age- and sex-matched healthy controls were selected randomly from the general population in 2015–2017. The healthy controls were unrelated to the patients. The patients and control groups were all from the Dravidian Kannada-speaking population of South India.

The inclusion criteria for patients with aSAH were diagnosis of aneurysmal SAH with the presence of symptoms suggestive of aSAH combined with subarachnoid blood found with the CT scan and a proven aneurysm found via conventional angiography. The exclusion criteria for selecting patients were: the presence of neuropsychiatric conditions like dementia, Parkinson's disease, epilepsy and psychoses; or if SAH resulted from a mycotic aneurysm, arterio-venous malformation or trauma. Demographic and clinical details were collected directly from the patients using structured questionnaires and data from the medical records section.

The inclusion criteria for healthy controls were: the absence of aneurysm, checked with digital subtraction angiography; similar demographic characteristics to the patient group (age, sex, ethnicity and dietary habits); no medical history of hemorrhage and no family history of aSAH in first-degree relatives.

The study protocol was approved by the Institute of ethics committee for human studies, NIMHANS, Bangalore. Written informed consent was obtained from all the participants.

DNA extraction and genotyping

Blood samples (5 ml) were collected from all the participants. Genomic DNA was isolated from the blood using the commercially available Machery-Nagel (MN) kit according to

the manufacturer's protocol. The purity and quantity of DNA was analyzed using a Nanodrop ND2000c spectrophotometer. DNA with a purity of 1.75–1.85 was used for genotyping analyses.

Genotyping of rs7412 and rs429358 was performed using a Taqman allelic discrimination assay (Applied Biosystems) with a commercially available primer probe set (assay ID C_904973_10, C_3084793_20). Genotyping was performed in duplicates using an Applied Biosystem7500 Fast Real-Time Cycler.

Statistical analysis

R.3.0.11 statistical software was used to statistically analyze the data. The continuous variables were expressed as means \pm SD and categorical variables as absolute values and percentages. The demographic characteristics of the patients and controls were compared using the χ^2 test for all categorical variables. Differences in genotype and allele frequencies between groups were analyzed using the χ^2 test. Association between *APOE* genotypes or alleles and aSAH risk were expressed as the odds ratio (OR) with 95% confidence intervals (CI), adjusted for the confounding effects of smoking, hypertension, drinking and diabetes mellitus using the logistic regression model. The Hardy–Weinberg equilibrium calculation was performed using an online tool at <http://www.oege.org/software/hwe-mr-calc.shtml>. $p < 0.05$ was considered statistically significant.

Results

Study population characteristics

The characteristics of patients with aSAH and healthy controls were shown in Table 1. Clinical data were available for all patients and healthy control subjects and there was no statistical significance in gender difference. The mean age was slightly higher in aSAH patients than in the controls with a p value of 0.170. Smoking and alcohol consumption were slightly more common in patients. The frequencies of hypertension and diabetes mellitus were also slightly higher in patients. The majority of the patients had aSAH with an ACOM (anterior communicating artery) aneurysm (43%) with a size of less than 15 mm (79.5%) and a WFNS grade of I (46%).

APOE polymorphism and risk of aSAH

The distribution of the *APOE* genotype and allele frequencies are shown in Table 2. The distribution of genotype frequencies of the control group was in Hardy–Weinberg equilibrium (rs7412; $p = 0.36$, rs429358; $p = 0.93$). The *APOE* allele frequencies in the controls ($n = 253$; $\epsilon_2 = 0.06$; $\epsilon_3 = 0.83$; $\epsilon_4 = 0.12$) were similar to the data obtained from a study carried out by Das et al. in a South Indian population ($n = 620$; $\epsilon_2 = 0.05$; $\epsilon_3 = 0.84$; $\epsilon_4 = 0.11$; $\chi^2 = 0.01$; $df = 2$; $p = 0.91$) [22]. In our study, there was no significant difference in *APOE* genotypes ($\chi^2 = 0.05$; $df = 5$; $p = 1$) and allele frequencies ($\chi^2 = 0.009$; $df = 2$; $p = 0.99$) between the patients and the controls.

A comparison of different genotype models of aSAH with healthy controls showed no significant difference in the data. The results of logistic regression analyses are shown in Table 3. A statistically significant association was found in the genotype model ϵ_3/ϵ_3 vs. ϵ_3/ϵ_4 (OR = 1.91, 95% CI = 1.16–3.14, $p = 0.01$). The comparisons of allele frequencies ϵ_2 vs. ϵ_3 (OR = 1.08, 95% CI = 0.61–1.91, $p = 0.78$), ϵ_2 vs. ϵ_4 (OR = 0.73, 95% CI = 0.36–1.46,

Table 1 Demographic and clinical characteristics of patients with aSAH and healthy controls

Characteristics	aSAH	Controls	<i>P</i>
Total no.	200	253	–
Sex (male/female)	77/123	114/139	0.160
Female (%)	61.50	69.50	–
Male (%)	38.50	57.00	–
Age (years)	50.72 ± 10.76	45.57 ± 17.73	0.170
Hypertension (yes/no)	109/91	105/148	0.006
Smoking (yes/no)	58/142	25/228	< 0.0001
Drinking (yes/no)	57/143	26/227	< 0.0001
Diabetes mellitus	71/129	60/193	0.005
Site of aneurysm (no./%)			
ACOM	86/43	–	
PCOM	12/6	–	
ICA	36/18	–	
MCA	37/18.5	–	
Multiple	22/11	–	
Basilar top	7/3.5	–	
Size of aneurysm (no./%)			
Small (< 15 mm)	159/79.5	–	
Large (15–25 mm)	37/18.5	–	
Giant (> 25 mm)	4/2	–	
WFNS Grade (no./%)			
Grade I	91/45.5	–	
Grade II	42/21	–	
Grade III	51/25.5	–	
Grade IV	16/8	–	

$p = 0.73$) and $\epsilon 4$ vs. $\epsilon 3$ (OR = 0.67, 95% CI = 0.43–1.05, $p = 0.008$) did not show any statistical significance.

When the aneurysms were classified according to location, size and WFNS grade and compared with different *APOE* genotype models and allele frequencies, only the PCOM aneurysm was statistically significant with $\epsilon 2$ vs. $\epsilon 3$ allele frequency (OR = 3.59, 95% CI = 1.11–11.64, $p = 0.03$). Classification of aneurysms according to *APOE* genotype frequency is shown in Table 4. Similarly, we performed comparisons between male vs. female, hypertensive vs. non-hypertensive, and diabetic vs. non-diabetic patients with different *APOE* genotype model and allele frequencies. None of the comparisons showed statistical significance with the *APOE* allele or genotype model.

Table 2 Distribution of *APOE* genotypes and alleles in patients with aneurysmal SAH and in control subjects

	Genotypes						Allele frequency		
	$\epsilon 2/\epsilon 2$	$\epsilon 2/\epsilon 3$	$\epsilon 2/\epsilon 4$	$\epsilon 3/\epsilon 3$	$\epsilon 3/\epsilon 4$	$\epsilon 4/\epsilon 4$	$\epsilon 2$	$\epsilon 3$	$\epsilon 4$
Patients	2 (1.0)	18 (9)	1	134 (67)	45 (22.5)	0	23 (5.75)	331 (82.75)	46 (11.5)
Controls	4 (1.60)	15 (5.92)	5 (1.97)	194 (76.6)	34 (13.4)	1 (0.4)	28 (5.53)	437 (86.36)	41 (8.1)

Numbers in parentheses are percentages

Table 3 Distribution of *APOE* genotypes and alleles in patients with aneurysmal SAH and control subjects

Genotypes	OR (95% CI)	P
ε2/ε2 vs. ε3/ε3	0.72 (0.13–4.01)	0.710
ε2/ε2 vs. ε3/ε4	2.64 (0.45–15.3)	0.270
ε3/ε3 vs. ε4/ε4	0.48 (0.01–11.92)	0.650
ε2/ε2 vs. ε2/ε3	2.4 (0.38–14.96)	0.340
ε2/ε2 vs. ε2/ε4	0.4 (0.02–6.17)	0.510
ε3/ε3 vs. ε2/ε3	1.73 (0.84–3.56)	0.130
ε3/ε3 vs. ε2/ε4	0.28 (0.03–2.5)	0.260
ε3/ε3 vs. ε3/ε4	1.91 (1.16–3.14)	0.01
ε4/ε4 vs. ε2/ε3	3.58 (0.13–94.31)	0.440
ε4/ε4 vs. ε2/ε4	0.81 (0.02–32.26)	0.910
ε4/ε4 vs. ε3/ε4	3.95 (0.15–100.13)	0.400
Alleles		
ε2 vs. ε3	1.08 (0.61–1.91)	0.780
ε2 vs. ε4	0.73 (0.36–1.46)	0.730
ε3 vs. ε4	0.67 (0.43–1.05)	0.080

Discussion

Subarachnoid hemorrhage caused by the rupture of an aneurysm is a condition with a low chance of recovery and a strong chance of life-long locomotor disability [23]. Guiding physicians in predicting the occurrence and rupture of an aneurysm will help them to save patients' lives and their capability to be productive. Various genetic polymorphisms are associated with the risk of developing aSAH. Apolipoprotein E polymorphism has emerged as one of the major genetic factors associated with the risk and prognosis of many neurological disorders and of hemorrhagic stroke in various populations.

Japan has the highest incidence rate of aSAH in the Asian population. One explanation for this higher incidence is the relatively high life expectancy in Japan [24]. Statistical reports state that the Japanese population has the highest median age (43 years old) [25]. The reported incidence rate of aneurysm in India ranges from 0.75–10.3%, and the North West Indian population, (lowest median age-28 years old) only has an incidence rate of 1% [26]. Therefore, age can be considered a risk factor for aSAH.

Another independent risk factor for aSAH is being female [23]. In our study, incidence of aSAH was 1.6 times higher in the female population than in the male population. The mean age of the female and male population was 55 and 40. The reason for the higher incidence rates in women are not still clear. Previously, it was observed that incident rate of aSAH was higher after menopause [27]. After menopause, a drop in sex hormones occurs [28], especially in estrogen. It has also been reported that estrogen has a protective role for SAH [29]. Estrogen has been reported to improve the lipid profile, reducing the risk of atherosclerosis [30, 31], which has been considered an important pathogenic mechanism for the formation of an aneurysm [32].

De Rooij et al. reported that at a younger age, the incidence of aSAH was higher in men than women [1]. But in our study, male and female subjects of less than 50 years of age were equally affected. There were 62.5% females who had multiple aneurysms in

Table 4 Distribution of APOE genotypes and allele frequencies according to aSAH subtypes

Variable	Case	ε 2/ ε 2	P	ε 2/ ε 3	P	ε 2/ ε 4	P	ε 3/ ε 3	P	ε 3/ ε 4	P	ε 4/ ε 4	P	ε 2 Vs ε 3 (P)	ε 4 Vs ε 3 (P)	
Total	200	2	18	1	134	45	0									
Site of Aneurysm																
ACOM	86	0	0.62	7	0.86	1	0.69	57	0.89	21	0.78	0	0.69	0.61	0.57	
PCOM	12	1	0.09	2	0.42	0	0.15	5	0.26	4	0.51	0	0.15	0.03	0.31	
ICA	36	0	0.82	2	0.92	0	0.33	30	0.61	4	0.18	0	0.33	0.26	0.12	
MCA	37	1	0.42	4	0.75	0	0.44	24	0.82	8	0.92	0	0.44	0.44	0.65	
Multiple	22	0	0.71	3	0.53	0	0.29	13	0.73	6	0.81	0	0.29	0.74	0.65	
Basilar top	7	0	0.29	0	0.82	0	0.11	5	0.91	2	0.69	0	0.11	0.69	0.81	
Size of Aneurysm																
Small (<15 mm)	159	1	0.70	14	0.95	1	0.87	109	0.89	34	0.83	0	0.89	0.81	0.81	
Large (15-25 mm)	37	1	0.50	3	0.86	0	0.45	22	0.68	11	0.46	0	0.45	0.67	0.39	
Giant (>25 mm)	4	0	0.17	1	0.37	0	0.06	3	0.88	0	0.95	0	0.06	0.50	0.61	
WFNS Grade																
Grade I	91	1	1	10	0.62	1	0.73	57	0.73	22	0.80	0	0.73	0.49	0.64	
Grade II	42	0	0.99	6	0.35	0	0.42	30	0.80	5	0.20	0	0.42	0.68	0.16	
Grade III	51	1	0.56	0	0.11	0	0.48	36	0.83	13	0.77	0	0.48	0.14	0.77	
Grade IV	16	0	0.57	2	0.67	0	0.16	11	0.94	5	0.54	0	0.16	0.99	0.67	
Male	77	1	0.50	10	0.37	1	0.62	51	0.95	14	0.52	0	0.62	0.27	0.62	
Female	123	1	0.85	8	0.46	0	0.70	83	0.96	31	0.66	0	0.81	0.35	0.76	
Hypertension (+)	109	1	0.94	9	0.83	0	0.75	74	0.94	25	0.94	0	0.75	0.71	0.96	
Diabetes mellitus (+)	71	0	0.82	5	0.63	1	0.46	51	0.74	14	0.69	0	0.55	0.30	0.57	
Alcohol (+)	57	1	0.64	10	0.11	1	0.37	33	0.54	12	0.85	0	0.55	0.04	0.85	
Smoking (+)	58	0	0.80	8	0.34	1	0.38	38	0.92	11	0.64	0	0.52	0.44	0.78	

our study, which concurs with previous findings that women are more likely to have multiple aneurysms than men [33].

APOE polymorphism has been associated with the risk of developing many central nervous system disorders, like Alzheimer's disease [34–36], vascular dementia [37], multiple sclerosis [38], cerebral infraction [39] and Parkinson's disease [40]. Many investigations have reported a positive association of *APOE* polymorphism with a risk of aSAH in different populations.

Liu et al. showed that $\epsilon 2/\epsilon 2$ and $\epsilon 2/\epsilon 3$ genotype frequencies were higher in patients with intracranial aneurysm in the Chinese population [41]. Tang et al. noted that $\epsilon 4$ allele carriers have unfavorable outcomes after aSAH in the Chinese population [42].

In the Japanese population, Kokubo et al. found that $\epsilon 4$ allele carriers have a 2.5-fold higher risk of aSAH [43]. Mineharu et al. suggested that the three alleles of *APOE* did not have any association with aSAH in Western Japan [44].

In the Caucasian population, McCarron et al. reported that the $\epsilon 2$ allele was significantly associated with the risk of different intracranial hemorrhage [13]. Kaushal et al. and Fontanella et al. found that $\epsilon 2$ and $\epsilon 4$ allele were not significantly associated with the risk of aSAH in United States and Italian population [45, 46].

A meta-analysis of nine case control studies found that in the Asian population, $\epsilon 2/\epsilon 2$ vs. $\epsilon 3/\epsilon 3$, $\epsilon 2/\epsilon 3$ vs. $\epsilon 3/\epsilon 3$, $\epsilon 2$ vs. $\epsilon 3$ and $\epsilon 2$ allele frequency were associated with the risk of aSAH. However, the study concluded that only the $\epsilon 2/\epsilon 2$ vs. $\epsilon 3/\epsilon 3$ genetic model was associated with the risk of aSAH in the Caucasian population [21].

In terms of lipoprotein metabolism, the main difference between *APOE3* and *APOE4* isoforms was that *APOE4* has greater affinity for very low-density lipoprotein (VLDL) receptor [47]. Therefore, *APOE4* impairs lipolytic processing and leads to the accumulation of VLDL in plasma [48]. The reason for pro-atherogenic lipoprotein–cholesterol distribution in the plasma of the *APOE4* isoform was because of the Cys112Arg substitution [49]. The presence of Arg112 in the *APOE4* isoform led to an altered interaction between the LDL receptor-binding domain and lipid-binding domain region, which was the reason for the higher binding affinity of this isoform to VLDL [50]. In ischemic stroke patients, the $\epsilon 3/\epsilon 4$ genotype was associated with elevated levels of very low-density lipoprotein and triglycerides [22].

Kokubo et al. reported 30% $\epsilon 3/\epsilon 4$ genotype frequency among aSAH patients in the Japanese population [43], but in the Chinese population, the $\epsilon 3/\epsilon 4$ genotype frequency was found to be 16% [41].

In our study, the $\epsilon 3/\epsilon 4$ genotype showed significant association with the risk of aSAH. The frequency for the $\epsilon 3/\epsilon 4$ genotype was higher in our patient group (23%) than the controls (14%). We also found that $\epsilon 4$ allele frequency was higher in the patient population, but we did not find any statistical significance.

One of the proposed mechanisms for aneurysm formation and progression is atherosclerosis [51]. It was reported that elevated levels of triglycerides and very low-density lipoprotein promote atherosclerosis [52]. $\epsilon 4$ carriers ($\epsilon 3/\epsilon 4$, $\epsilon 4/\epsilon 4$, $\epsilon 2/\epsilon 4$ genotype) have been associated significantly with risk for atherosclerosis and elevated levels of very low-density lipoprotein [53, 54]. From all these findings, it can be summarized that the $\epsilon 3/\epsilon 4$ genotype is one of the risk factor for aSAH, since it can promote atherosclerosis.

In this study, of the aSAH subtypes, the $\epsilon 2$ allele has a significant association with PCOM aneurysm. The PCOM aneurysm is the third most common Circle of Willis

aneurysm. PCOM arteries are present at the base of the brain and form a part of the Circle of Willis [55]. It was reported that the $\epsilon 2$ allele is associated with an elevated concentration of plasma triglycerides [56] and that $\epsilon 2/\epsilon 3$ genotype carriers were less likely to survive a stroke [57]. Also, the presence of the *APOE2* allele may indicate susceptibility to the development of fibrinoid necrosis and microaneurysm [58].

The main difference between *APOE3* and *APOE2* isoform was that the latter rarely binds to low-density lipoprotein receptors [59]. *APOE2* differs from the *APOE3* isoform by Arg to Cys amino acid substitution at position 158, which is close to the LDL receptor-binding domain region [60]. Cys158 disrupts the natural salt bridge between Asp152 and Arg154 in the LDLR recognition site, which impairs the binding to the LDL receptor [61]. This makes the *APOE2* isoform unable to promote the clearance of low density lipoprotein and triglycerides from the plasma [62]. Bolger et al. noted that one of the reasons for saccular aneurysmal disease is the elevated serum level of low-density lipoprotein [63]. This suggests that the $\epsilon 2$ allele has a role in elevating the levels of low-density lipoprotein and thereby can be a causal factor in aSAH.

Conclusion

APOE polymorphism can be associated with the risk of aSAH in the South Indian population. When compared with the site of aneurysm, the $\epsilon 2$ allele was found to be associated with PCOM aneurysm. Further comprehensive studies are required to confirm these findings.

Abbreviations

ACOM: Anterior communicating artery; *APOE*: Apolipoprotein E; aSAH: Aneurysmal subarachnoid hemorrhage; CI: Confidence interval; ICA: Internal carotid artery; LDLR: Low-density lipoprotein receptor; MCA: Middle cerebral artery; OR: Odds ratio; PCOM: Posterior communicating artery; VLDL: Very low-density lipoprotein; WFNS: World Federation of Neurological Surgeons; χ^2 test: Chi square test

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Availability of data and materials

All the data analyzed during this study are included in this article. Additional information related to this study is available from the author for correspondence upon reasonable request.

Authors' contributions

AS performed sample collection, DNA extraction and genotyping, co-conceived the study and participated in its design, acquired data, interpreted the results, and drafted and revised the manuscript. SMK participated in the design of the study, helped in the interpretation of results, and performed statistical analyses. DIB, NR and WV made theoretical contributions and approved the version of the manuscript to be published. CGK co-conceived the study, helped in study design, contributed to the review of manuscript, and gave the final approval to publish. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The study protocol was approved by the Institute of Ethics Committee for Human Studies, NIMHANS, Bangalore, India (Item No. III, Sl.No.3.02, Basic Sciences). Written informed consent was obtained from all the participants.

Consent for publication

Written informed consent for publication of data was obtained from all the participants.

Competing interests

The authors declare that no competing interests exist on the materials or methods used in this study and findings specified in this paper.

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